# In Vivo Antidiabetic Properties of Etlingera elatior Leaf Extract in Alloxan-Induced Diabetic Rats

By Dora Dayu Rahma Turista Qurrotu 'yunin Lathifah

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#### In Vivo Antidiabetic Properties of Etlingera elatior Leaf Extract in Alloxan-Induced Diabetic Rats

Dora Dayu Rahma Turista<sup>1\*</sup>, Qurrotu A'yunin Lathifah<sup>2</sup>, Arif Nur Muhammad Ansori<sup>3</sup>, Yulanda Antonius<sup>4</sup>, Gabrielle Ann Villar Posa<sup>5</sup>, Wahyu Choirur Rizky<sup>6</sup>, Tim Godefridus Antonius Dings<sup>7</sup>, Galiya Kazhibayeva<sup>8</sup>, Karina Omarova<sup>8</sup>, Irina Anikina<sup>8</sup>

<sup>1</sup>Educational Biology Department, Faculty of Teacher Training and Education, Mulawarman University, Samarinda, Indonesia.

<sup>2</sup>Department of Medical Laboratory Technology, STIKES Hutama Abdi Husada, Tulungagung, Indonesia.

<sup>3</sup>Doctoral Program in Veterinary Science, Faculty of Veterinary Medicine, Universitas Airlangga, Surabaya, Indonesia

Faculty 25 Biotechnology, University of Surabaya, Surabaya, Indonesia.

<sup>5</sup>School of Environmental Science and Management, University of the Philippines Los Baños, Los Baños, Philipines.

<sup>6</sup>College of Medicine, Sulaiman Al Rajhi University, Al Bukayriyah, Qassim, Saudi Arabia.

<sup>7</sup>College of Medicine, Maastricht University, Maastricht, The Netherlands.

\*Corresponding author:

pucational Biology Department,

Faculty of Teacher Training and Education,

Mulawarman University,

Samarinda, East Borneo 75119, Indonesia

E-mail address: doraturistaofficial@gmail.com

Tel: +6285730477725

#### ABSTRACT:

Diabetes mellitus is a metabolic disease characterized by hyperglycemia. Application of alloxan in experimental animals can cause Diabetes mellitus. The secondary metabolites of *Etlingera elatior* can be used as raw materials for diabetes mellitus drug. This study aims to determine the antidiabetic potential of ethanol extract of *Etlingera elatior* leaves by *in vivo* study. A total of 32 rats were divided into 6 groups, namely NC, DC, PC, DE1, DE2, and DE3. The results of data analysis using multivariate ANOVA on blood glucose level data every week showed p (0.000) < (0.05), and the results of data analysis using one way ANOVA on pancreatic  $\beta$  cell count data also showed that p (0.000) < (0.05). *Etlingera elatior* leaf ethanol extract has antidiabetic activity since it could reduce blood glucose levels and increase the number of pancreatic  $\beta$  cells through several mechanisms. The mechanism is triggered by phytochemical compounds contained in the leaf extract of *Etlingera elatior*.

**KEYWORDS:** Alloxan, diabetes mellitus, *Etlingera elatior*, blood glucose level, pancreatic β cells

#### 12 INTRODUCTION:

Diabetes mellitus is metabolic diseases that is characterized mainly by hyperglycemia which caused by defect of insulin secretion, action of insulin, 14 both. Chronic hyperglycemia in diabetes resulting a long-term complication. This disease is related to obesity, hypertension, and abnormal lipid profile, such as high triglyceride levels, low of high-density lipoprotein (HDL), high total cholesterol<sup>2</sup> and increases the risk of cardiovascular diseases<sup>3,4</sup>. People with diabetes mellitus reached 422 million worldwide and caused 1.6 million deaths 59 h year<sup>5</sup>. The morbidity rate of diabetes continuously rising until now. The world has agreed that it will stop the increase in the number of diabetes cases by 2025<sup>5</sup>.

<sup>&</sup>lt;sup>8</sup>Department of Biotechnology, Toraighyrov University, Pavlodar, Kazakhstan.

According to American Diabetic Association (ADA, 2021) Diabetes mellitus is divided into 3 types, namely type 1 diabetes mellitus (T1DM), type 2 diabetes mellitus (T2DM), and other specific types. T1DM is caused by destruction to the  $\beta$ -cells in the pancreas which cannot prod 18 insulin optimally. It is either causing by immune-mediated process or idiopathic. Moreover, T2DM is caused by insulin resistance with relative insulin deficiency to a predominantly secretory defect. Other specific types of diabetes can be cau 28 by genetic causes, diseased pancreatic exocrine, endocrinopathies, drug- or chemical-induced, or infections. Insulin regulates blood glucose levels by promoting glucose uptake from the blood to the cells. The pancre 12 an organ that synthesizes and secretes the insulin, specifically by the  $\beta$  cells of the Langerhans islets. Insulin secreted by the pancreas into the portal vein then enters to the liver. Furthermore, it is distributed throughout the body through the blood circulation. The disorders of the Langerhans islets cells have an impact on insulin secretion. When insulin secretion is impaired, the blood glucose 26 eased, and it cannot even enter to the cells. Consequently, cells lack glucose as an energy-forming number 15 rial.

Alloxan is a diabetogenic agent that commonly used to induce diabetes in experimental animals. It is an organic compound, urea derivative, and glucose analog which has carcinogenic and cytotoxic effect. It is chemically known as 5,5-dihydroxy pyrimidine-2,4,6-trione<sup>6,7</sup>. The use of alloxan in experimental animals can cause type 1 diabetes<sup>8</sup>. Alloxan works by inhibiting the glucokinase enzyme and inducing the formation of ROS. The glucokinase is a glucose sensor of beta cells. Furthermore, the inhibition of glucokinase inhibited the insulin 36 etion by  $\beta$ -cell, whereas ROS induce necrosis of  $\beta$  cells resulting in insulin-dependent diabetes<sup>6</sup>. During 15 ours after administration of alloxan at a dose of 170 and 200 mg/kg BW to the experimental animals showed several phases of glucose response were observed<sup>8,9</sup>. Intravenous alloxan-induced rats show biochemical changes in the blood <sup>10</sup>.

Plants produced secondary metabolites that can be used as a raw material of drug<sup>6</sup>, such as *Etlingera elatior*. The herbs medicines is commonly made from secondary metabolites and it showed various biological effects<sup>11</sup>. *E. elatior* contains a secondary compounds of alkaloids, terpenoids, steroids, saponins, and flavonoids<sup>7</sup>. Flavonoids and alkaloids can trigger the regeneration of pancreatic  $\beta$ -cells<sup>8,9</sup>. Flavonoids can also protect the pancreatic  $\beta$ -cells and help them to survive<sup>12</sup>. The flavonoids 21 so act as antioxidants and it can increase insulin secretion<sup>11</sup>. Furthermore, it could repair the amaged pancreatic  $\beta$ -cells, and improve the insulin secretion. Therefore, the blood glucose level could be stable. This study aimed to determine antidiabe 41 otential of the ethanol extract of *Etlingera elatior* leaves by *in vivo* study. In brief, it could be determined through blood glucose levels and the number of pancreatic  $\beta$  cells in rats which injected by alloxan.

#### MATERIAL AND METHODS:

#### Material selection:

Alloxan monohydrate is procured from Merck (Sigma Aldrich). The standard drug Glibenclamide and Carboxymethylcellulose (CMC) were used as a negative control. Furthermore, the leaves of *Etlingera elatior* from Zingiberaceae family was used for the experiment. Furthermore, ethanol was selected as solvent for extraction.

#### Extract preparation:

The leaves of *Etlingera elatior* were washed, cut, and air-dried. The dried leaves was then processed to be powder. Furthermore, 50 grams of *Etlingera elatior* powder was macerated with 250 mL of 96% ethanol for 48 hours at room temperature. Furthermore, solution of leaf extract was filtered and the filtrate was concentrated using a rotary evaporator<sup>13</sup>.

#### **Extract screening:**

Screening of Etlingera elatior leaf extracts was conducted by following the Harbour Method 14:

Flavonoids: 1 ml of concentrated extract was put into a test tube, about 1-2 ml of hot methanol was added, then the Mg metal powder was added. Furthermore, about 0.5 ml of concentrated HCl was added. If it produced a red or orange color, then the extract was positive for flavonoids.

Tannins: About 1.5 ml of concentrated extract was put into a test tube, then a few drops of hot distilled water were added. Moreover, it was cooled and filtered. Furthermore, three drops of 10% NaCl were filtered. Then two drops of FeCl<sub>3</sub> were added. If it produced a blackish green/dark blue color, then the sample was positive for tannins.

Saponin (forth method): One ml of concentrated extract was put into a reaction tube, then 5 ml of distilled water was added and shaken for 30 seconds. If it caused foam and did not disappear for 30 seconds, then the extract would be positive for saponins. However, to maintain the bias foam, 1 M HCl was added.

Phenolic: 1 ml of concentrated extract was put into a test tube, then 10 drops of 1% FeCl<sub>3</sub> were added. The extracts was considered contained of phenol if it produced green, red, purple, blue, or solid black color.

Alkaloids: 1 ml of concentrated extract was put into a test tube, then 3-5 drops of Dragendroff's reagent were added. The positive reaction occured when a brown or orange precipitate was formed.

Terpenoid/Steroid: 1 ml of concentrated extract was put into a test tube, then 0.5 ml of chloroform was added. Moreover, 0.5 ml of anhydrous acetate was added. The mixture was then added with 3-5 drops of concentrated H<sub>2</sub>SO<sub>4</sub>

through the wall. If a green or blue color was formed, then the extract was considered positive for steroids. Meanwhile, if a purple or brown ring was formed, the extract was determined as positive for triterpenoids.

#### Animal preparation:

This study was approved by the ethics committee for animal research at Faculty of Veterinary Medicine, Universitas Airlangga (2.KE.041.04.2020). In this study, the experimental were used male rats (*Rattus norwegicus*) with Wistar strain. Rats were at age 2-3 months and weighed about 175-200 grams with good health conditions.

#### Induction of diabetes mellitus:

The rats were acclimated in the laboratory for 7 days and ensured that their blood glucose levels were below 200 mg/dL before alloxan administration. White rats were induced to be diabetic by injecting a single dose of alloxan 175 mg/kg body weight (BW) intraperitoneally<sup>15</sup>.

#### Experimental design:

The dose of *Etlingera elatior* leaf extract was using a human dose of 10 grams/50 kg BW or 14 grams/70 kg BW. Hence, it was converted into rats with weight 200 grams. In detail, it was calculated as follow: 0.018 x 14 grams = 0.252 grams or 252 mg. The *E. elatior* leaf extract was injected once a day according to the group dose with a treatment duration of 21 days. The determination of the number of rats was calculated using the Frederer formula. Total of 32 rats were divided into 6 groups, such as:

NC: Normal Control (treated with CMC in distilled water and non-alloxan-induced)

DC: Diabetes mellitus Control (CMC-treated in alloxan-induced)

PC: Positive Control (Glibenclamide-treated in allox an-induced)

DE1: Diabetic Extract 1 (200 mg ethanol leaf extract/200 gram BW rats in alloxan-induced)

DE2: Diabetic Extract 2 (250 mg ethanol leaf extract/200 gram BW rats in alloxan-induced)

DE3: Diabetic Extract 3 (300 mg ethanol leaf extract/200 gram BW rats in alloxan-induced)

#### Research data collection:

Blood glucose levels were measured every 7 days, i.e. days 0, 7, 14, 21, and 28 after treatment. Blood was taken from the tail and it was examined by using a glucometer. The examination of pancreatic cell structure was carried out 28 days after treatment. The rats were dissected and their pancreas was taken for histopathological preparations by using the Hematoxylin Eosin (HE) staining method. After staining, the preparations were examined using a microscope with a magnification of 400× within 5 fields of view.

#### Data analysis:

Data analysis using the IDB SPSS software (version 20). Weekly blood glucose levels were analyzed by multivariate ANOVA and the number of pancreatic  $\beta$ -cells were analyzed by one way ANOVA. Furthermore, the post hoc was analyzed by using the LSD test. The P-value was <0.05 (p<0.05) based on statistical significance.

#### **RESULTS AND DISCUSSION:**

Plants contain a phytochemical compounds which resulted from secondary metab 45 n. The results of a qualitative screening of the phytochemical content of the *Etlingera elatior* leaf ethanol extract is presented in Table 1.

Table 1. Phytochemical content of Etlingera elatior leaves ethanol extract

Phytochemicals	Etlingera elatior Leaf
Flavonoid	+
Tannin	+
Saponin	+
Phenolic	+
Alkaloid	+
Triterpenoid	+
Steroid	-

This experiment was conducted after the rats blood glucose levels reached above 200 mg/dL due to alloxan induction. In general, this study resulted in two data which are presented in the form of mean, namely blood glucose levels every the horizontal peak from week 0 to week 4. Moreover, the number of pancred  $\beta$  cells after the rats were dissected at week 4. Blood glucose levels per week are presented in Figure 1 and the number of pancreatic  $\beta$  cells are presented in Figure 2.

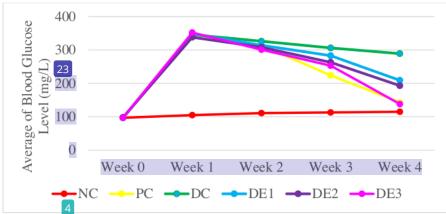


Figure 1. Rats Blood Glucose Levels

The data of rats' blood glucose levels in every week were carried out by comparative analysis using multivariate ANOVA and it was found that p (0.000) < (0.05). Furthermore LSD test was carried out and it was found that at week 4, there was no significant difference observed for DE3 as compared to NC (p=0.054) and PC (p=0.607).

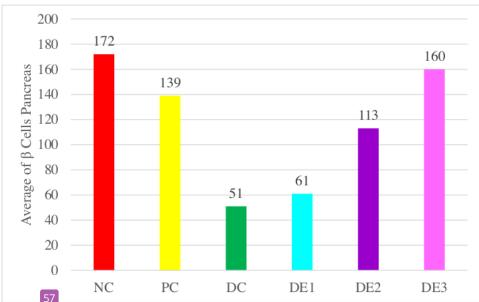


Figure 2. The Number of Rat Pancreatic Beta Cells

The 24 a on the number of pancreatic β cells in rats was carried out by a comparative analysis using one way ANOVA and the results showed a significant difference between groups (p<0.05). Furthermore, the LSD test was carried out and it was found that DE3 showed no difference to NC (p: 0.580) and PC (p: 0.307).

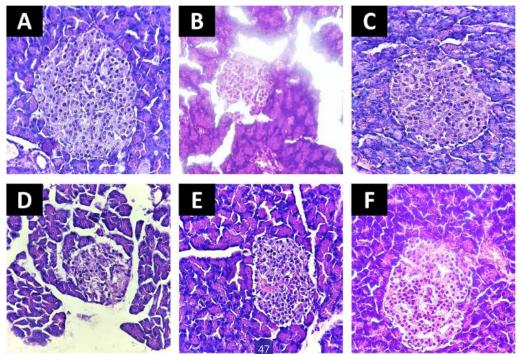


Figure 3. Photomicrograph of Rats Pancreas Histopathology stained with Hematoxylin Eosin in 400× magnification: A) Normal Control: Normal rats that given CMC, B) Diabetes mellitus Control: Diabetes mellitus rats that just given CMC, C) Positive Control: Diabetes mellitus rats that given Glibenclamide, D) Diabetic Extract 1: Diabetes mellitus Rats that given 200 mg ethanol leaf extract/200 gram BW rats, E) Diabetic Extract 2: Diabetes mellitus Rats that given 200 mg ethanol leaf extract/200 gram BW rats, F) Diabetic Extract 3: Diabetes mellitus Rats that given 300 mg ethanol leaf extract/200 gram BW rats.

Figure 3A is histopathology of the pancreas of normal rats without alloxan induction. It showed that the islets of Langerhans are in good condition with a large number of  $\beta$ -cells without damage observed. Figure 3B is the histopathology of diabetes mellitus rats due to alloxan induction and without therapy. It appeared that the islets of Langerhans are damaged, the boundaries of the islets are not clear, and the  $\beta$ -cells are few due to necrosis or apoptosis. Figure 3C is the histopathology of the diabetic rats pancreas that was treated with glibenclamide for 4 weeks, it appears that the islets of Langerhans are good with a large number of  $\beta$  cells and there is no damage to  $\beta$  cells. Whilst, figure 3D is the histopathology of the diabetic rats' pancreas that was treated with 200 mg ethanol leaf extract/200 gram BW rats, it shows that islets of Langerhans are damaged and the  $\beta$  cells are few due to necrosis or apoptosis. Figure 3E is the histopathology of the diabetic rats' pancreas that was treated with 250 mg ethanol leaf extract/200 gram BW rats. It showed that the number of  $\beta$  cells are increased. Figure 3F is the histopathology of the diabetic rats' pancreas that was treated with 300 mg ethanol leaf extract/200 gram BW rats. It shows the best results with good Langerhans islets and the number of  $\beta$  cells under conditions similar to those of normal mouse pancreas and glibenclamide-treated rats' pancreas.

The blood glucose level of rats increased above the normal limit (200 mg/dL) after alloxan administration w 27 single dose of 175 mg/200g BW (Figure 1). This is because alloxan is a toxic 27 bstance that can cause damage to pancreatic β-cells. Alloxan is a highly toxic compound and it is commonly used to induce diabetes mellitus in animals. In brief, it works in two ways, such as causing insulin dependence and causing necrosis of pancreatic β-cells. Alloxan induced partial degradation of pancreatic cells so that the quality and 22 antity of insulin are impaired. Insulin synthesis begins with the formation of preproinsulin and it is processed into proinsulin and subsequently converted into insulin and C-61 tide is stored in secretory granules and secreted on demand.

Blood glucose level in diabetes mellitus rats is related to the condition and number of  $\beta$ -cells in the Langerhans is lets. In normal rats, the pancreas appeared normal w 17 arge is lets of Langerhans without any damage to  $\beta$ -cells (Figure 1a) and a high number of  $\beta$ -cells (Figure 2). The size of the pancreatic Langerhans is lets in diabetic rats is also smaller and many 19 heir  $\beta$ -cells are damaged (Figure 3b). Furthermore, the number of  $\beta$ -cells decreased (Figure 2). Is lets consisting mostly of only remnants of  $\alpha$  and  $\beta$  cells show decreased in the islet size,  $\beta$  granules, and granular endoplasmic reticulum<sup>17</sup>. This is due to the pathological effect of the alloxan.

Alloxan is very harmful to pancreatic  $\beta$ -cells which is insulin-producing cells but it is not harmful to  $\alpha$  cells as glucagon-producing cells<sup>15</sup>. The alloxan is selectively absorbed very quickly by pancreatic  $\beta$  cells since it has similarities to glucose in molecular form and hydrophilicity, thus eventually accumulates in the cells<sup>8</sup>. The transport of alloxan into the cytosol of  $\beta$ -cells is also similar to that of glucose, namely through facilitated diffusion by glucose transport protein 2 (GLUT2). The GLUT2 transporter is usually located on the plasma membrane of  $\beta$ -cells.

Alloxan is a highly unstable compound and it is susceptible to a redox reaction<sup>8</sup>. It caused diabetes through the inactivation of essential sulfhydryl enzymes which is involve the combination of essential sulfhydryl groups and oxidized to disulfide bonds and vice versa<sup>18</sup>. Alloxan reduction produces dialuric acid which is further oxidized by alloxan and the same step is continuously repeated. The autoxidation of dialuric acid is believed to be very important in the diabetogenic action of alloxan because it formed intracellular reactive oxygen species (ROS), such as hydroxyl radicals and superoxide radical anions that can trigger the tissue damage<sup>19</sup>. This reaction will occur in cycle and continue when there are intracellular thiols, especially glutathione (GSH)<sup>8</sup>.

Alloxan in 6 cs ROS activation which is a common mediator of necrotic cell death. The β-cells are highly susceptible to ROS. It will inhibit insulin or insulin-like growth factor (IGF)-1, insulin receptor (IR), 21 lin receptor substrate (IRS) -1, and phosphatidylinositol-3 kinase (PI3K)/Akt kinase. Furthermore, it induced the β-cell damage, decrea 37 insulin secretion and led to diabetes<sup>20</sup>. ROS caused mitochondrial injury, cell membrane damage, disturbing ion balance through protein damage, lipid peroxidation, and oxidative DNA damage<sup>21-23</sup>. Furthermore, ROS are associated with oxidative stress that causes damage to lipids, proteins, and DNA<sup>24,25</sup> which resulting in apoptosis cell control in addition to necrosis, the dead β-cell is also caused by apoptosis, so that the number of β-cells decreased<sup>28-30</sup>. Necrosis can induce the release of inflammatory cytokines, such as IL-1, IL-4, IL-13, and other inflammatory mediators. Whereas apoptosis can lead to the release of anti-inflammatory cytokines including TGF-β and IL-10<sup>31</sup>. Moreover, oxidative stress can cause diabetes-related cell and tissue (7 nage<sup>32,33</sup>).

Administration of ethanol extract of Etlingera elatior leaves could red 43 the blood glucose levels of diabetic rats (Figure 1). During 200 mg/200 g BW, the treatment showed that blood glucose levels 56 eased but it did not reach the normal levels. The dose of 250 mg/20 50 BW showed the decrease of normal blood glucose levels (>200 mg/dL) after week-4. The decreased of maximum blood glucose level was found in the diab 14 rats treated with 300 mg/200 g BW of extract. In addition, it demonst 11 d the highest level in this study. This is consistent with the results of the data analysis which showed the effect of blood glucose reduction in the diabetic rats. This effect is considered as dose-dependent. However, treatment with 300 mg/200 g BW is also showed that the state of the Langerhans islets is looked substantial (Figure 3d) with number of beta cells close to non-diabetic rats (Figure 2). This condition is related to the phytochemical content of extract. Treatment with higher dose increased the levels of phytochemicals absorbed by the rat's body.

Etlingera elatior leaf ethanol extract contains of compounds, such as flavonoids, tannins, saponins, phenolics, alkaloids, and triterpenoids (Table 1). The flavonoids are phenolic compounds resulting from secondary plant metabolism which are classified into subclasses flavonols, flavanones, flavones, isoflavones, flavonois, anthocyanidins, and flavanonols<sup>12,34,35</sup>. Flavonoids can prevent the diabetes and its complications<sup>36,37</sup>. Flavonoids can strengthen the capacity of insulin secretion and the process of  $\beta$ -cell survival through the antioxidant and antidiabetic 31 vities<sup>37</sup>.

Tannins are polyphenolic biomolecules that had high molecular weight and widely distributed in various plant species<sup>38</sup>. In detail, it divided into two groups, such as hydrolyzed tannins (molecules with polyhydroxy componen 40 and thick tannins (formed from the condensation of flavanols)<sup>38,39</sup>. Tannins also had an antidiabetic activity by inhibiting the activity of  $\alpha$ -amylase<sup>40</sup> and  $\alpha$ -glucosidase enzymes<sup>41</sup>.

Saponins are secondary metabolites of amphipathic gl 13 sides which are synthesized by various plant species and it has high molecular weights. Moreover, it consists of a sugar moieties, such as glucose, galactose, glucuronic acid, xylose, rhamnose or methylpentose, glycosides linked to the hydrophobic aglycones (sapogenins) which may be triterpenoids or steroids  $^{42,43}$ . Saponins with 55 sugar group are called monodesmosidic, but saponin with two sugar groups are called bidesmosidic  $^{44}$ . Saponins have been reported to stimulate insulin release, blocking the formation of glucose in the bloodstream  $^{45}$ , having mild inhibition of  $\alpha$ -amylase enzymes and inhibition of strong against the enzyme  $^{46}$ .

Alkaloids are derived from natural sources which contained of nitrogen atoms in heterocyclic compounds. Furthermore, it has various types of ring structures  $^{45,47}$ . Furthermore, alkaloids also h<sub>2</sub>38 hypoglycemic activity  $^{48}$ . The biomolecule is believed to have anti-diabetic properties since it can overcome the insulin resistance, reduce blood glucose levels, and accelerate the β-cell rejuvenation in diabetic experimental animals  $^{49}$ .

Triterpenes are produced by plants and marine animals which are formed through the composition of squalene epoxide followed by condensation, esters or glycosides (saponins) form in free condition<sup>45,50</sup>. Triterpen 10 re believed to have antidiabetic activity and inhibit diabetes the complications<sup>51,52</sup>. It involves in several signaling mechanisms including

activation of insulin signaling pathways, inhibition of PTP1B, GP, 11 $\beta$ -HSD1,  $\alpha$ -glucosidase,  $\alpha$ -amylase, and activation of AMPK and PPAR<sup>51</sup>.

Various results *in vitro* and *in vivo* showed that flavonoids<sup>32,53-59</sup>, tannins<sup>60–62</sup>, saponins<sup>63–68</sup>, phenolics<sup>69–75</sup>, alkaloids<sup>76–81</sup>, and triterpenoids<sup>51,52,76,82–89</sup> are phytochemical compound that have antioxidant activity. The endogenous and exogenous antioxidants had an essential role in cell defense mechanisms. It protects and repairs the cell damage by inhibiting the ROS production and scavenging free radicals<sup>32,33,90</sup>. Exogenous antioxidants derived from natural ingredients strengthen the endogenous antioxidant defenses<sup>91,92</sup>. The increased levels of antioxidants in the body could protect against degenerative diseases<sup>39,92</sup>.

The pancreas has a low levels of antioxidants. Therefore, *Etlingera elatior* leaf extract is considered to increase the antioxidant capacity through Nrf2 activation. Phytochemicals can activate Nrf2, thereby increasing antioxidant response and preventing β-cell death<sup>93</sup>. Previous studies shown that the ethanol extract of *Centipeda minima*<sup>94</sup>, with aqua 63 extracts of *Polygonatum sibiricum*<sup>95</sup>, and ethanol e 3 act of *Sargassum horneri* (Turner) C. Agardh<sup>96</sup>, could also activate the Nrf2 signaling pathway. Moreover, the Nuclear factor erythroid 2-related factor 2 (Nrf2) is a transcription factor that manage the cellular defense against toxic and oxidative atta 9 s through the gene's expression in oxidative stress response and drugs detoxification<sup>97</sup>. When the cells are exposed to oxidative stress or electrophilic compounds, the Nrf2 will dissociates from Keap1 and enters the nucleus to bind antioxidant-responsive elements in genes encoding antioxidant enzymes<sup>98</sup>.

Blood glucose levels are also related to the condition and number of pancreatic  $\beta$ -cells. Furthermore, accurate respond to the blood glucose levels with a sufficient number of  $\beta$ -cells are required 99,100. The recent study revealed that insulin-producing cells in mice could be regenerated 101.

The administration of *Etlingera elatior* leaf extract stimulates the  $\beta$ -cells regeneration, so that the number of  $\beta$ -cells increased. The regeneration potential of  $\beta$ -cells is very limited in the absence of external stimuli, but in the presence of external stimuli there is a sufficiently strong regenerative the  $\beta$ -cell mass expansion resulting from the activation of inactivated precursors/progenitors or stem cells<sup>102</sup>.

Etlingera elatior leaves have the potential to be used as herbal medicines for diabetes mellitus treatment because they have antidiabetic properties, and it is easy to obtain. Several phytochemicals from medicinal plants have been developed as new types of 17 betes mellitus therapy<sup>89</sup>. Plants are a source of natural antioxidants and effective herbal medicines related to its anti-diabetic compounds, such as 51 vonoids, tannins, phenolics, and alkaloids. These compounds enhanced the pancreatic tissue performance by increasing insulin secretion or decreasing the glucose absorption 5 the intestine 79. Moreover, the phytotherapy is excellent since it is safe, cheap, and abundantly available in nature. However, further research is needed to determine the mechanism of action and the molecular interactions of compounds within the body.

#### CONCLUSION:

Etlingera elatior leaf ethanol extract was able to significantly reducing the blood glucose levels and increasing the number of pancreatic  $\beta$ -cells. The Etlingera elatior leaf is cons 5 red as a potential candidate for antidiabetic mellitus drug since it is determined as safe, cheap, and easy to obtain. However, further research is needed to determine the mechanism of action and the molecular interaction of the compounds within the body.

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#### CONFLICT OF INTEREST:

The authors declare no conflict of interest.

#### REFERENCES:

- Shah NA, Khan MR. Antidiabetic effect of Sida cordata in alloxan induced diabetic rats. Biomed Res Int. 2014; 2014;67129. doi:10.1155/2014/671294
- Al Mansour MA. The prevalence and risk factors of type 2 diabetes mellitus (DMT2) in a semi-urban Saudi population. Int J Environ Res Public Health. 2020;17(1):1-8. doi:10.3390/ijerph17010007
- Matheus ASDM, Tannus LRM, Cobas RA, Palma CCS, Negrato CA, Gomes MDB. Impact of diabetes on cardiovascular disease: An update. Int J Hypertens. 2013; 2013:653789. doi:10.1155/2013/653789
- Schmidt AM. Diabetes Mellitus and Cardiovascular Disease. Arterioscler Thromb Vasc Biol. 2019;39(4):558-568. doi:10.1161/ATVBAHA.119.310961
- WHO. Diabetes. World Health Organization. Published 2021. Accessed March 21, 2021. https://www.who.int/health-topics/diabetes#tab=tab\_1
- Lenzen S. The mechanisms of alloxan- and streptozotocin-induced diabetes. Diabetologia. 2008;51(2):216-226. doi:10.1007/s00125-007-0886-7

- Nosiri CI, Atasie OC, Alvan LC, Ifedigbo O. Histopathology of the Pancreatic Cells of Alloxan Induced Wistar Rats Treated with Psidium Guajava Ethanolic Leaf Extract. IOSR J Biotechnol Biochem. 2016;2(4):28-32.
- Ighodaro OM, Adeosun AM, Akinloye OA. Alloxan-induced diabetes, a common model for evaluating the glycemic-control potential of therapeutic compounds and plants extracts in experimental studies. Med. 2017;53(6):365-374. doi:10.1016/j.medici.2018.02.001
- Yin P, Wang Y, Yang L, Sui J, Liu Y. Hypoglycemic Effects in Alloxan-Induced Diabetic Rats of the Phenolic Extract from Mongolian Rups Cups Enriched in Ellagic Acid, Kaempferol and Their Derivatives. Molecules. 2018;23(5):1-14. doi:10.3390/molecules23051046
- Lucchesi AN, Cassettari LL, Spadella CT. Alloxan-induced diabetes causes morphological and ultrastructural changes in rat liver that
  resemble the natural history of chronic fatty liver disease in humans. J Diabetes Res. 2015; 2015:494578. doi:10.1155/2015/494578
- A. Hussein R, A. El-Anssary A. Plants Secondary Metabolites: The Key Drivers of the Pharmacological Actions of Medicinal Plants. In: Herbal Medicine. IntechOpen; 2019:11-29. doi:10.5772/intechopen.76139
- Ghorbani A, Rashidi R, Shafiee-Nick R. Flavonoids for preserving pancreatic beta cell survival and function: A mechanistic review. Biomed Pharm. 15 her. 2019;111(October 2018):947-957. doi:10.1016/j.biopha.2018.12.127
- Gberikon GM, Adeoti I., Aondoaclaa. AD. Effect of ethanol and aqueous solutions as extraction solvents on phytochemical screening and antibacterial activity of fruit and stem bark extracts of Tetrapleura tetrapteraon, Streptococcus salivarus, and Streptococcus mutans. IntJCurrMicrobiolAppSci. 2015;4(5):404-410.
- 14. Harbome JB. Metode Fitokimia: Penuntun Cara Modern Menganalisis Tumbuhan. Second. Indonesia University; 1987.
- Federiuk IF, Casey HM, Quinn MJ, Wood MD, Ward WK. Induction of type-1 diabetes mellitus in laboratory rats by use of alloxan: Route of administration, pitfalls, and insulin treatment. Comp Med. 2004;54(3):252-257.
- Fu Z, R. Gilbert E, Liu D. Regulation of insulin synthesis and secretion and pancreatic beta-cell dysfunction in diabetes. Curr Diabetes Rev. 2012;9(1):25-53. doi:10.2174/15733998130104
- Muthuraman P, Senthilkumar R, Srikumar K. Alterations in beta-islets of Langerhans in alloxan-induced diabetic rats by marine Spirulina platensis. J Enzyme Inhib Med Chem. 2009;24(6):1253-1256. doi:10.3109/14756360902827240
- Patterson JW, Lazarow A, Levey S. Reactions of alloxan and dialuric acid with the sulfhydryl group. J Biol Chem. 1949;177(1):197-204. doi:10.1016/S0021-9258(18)57075-7
- Munday R. Dialuric acid autoxidation. Biochem Pharmacol. 1988;37(3):409-413. doi:10.1016/0006-2952(88)90207-9
- He L, He T, Farrar S, Ji L, Liu T, Ma X. Antioxidants maintain cellular redox homeostasis by elimination of reactive oxygen species. Cell Physiol Biochem. 2017;44(2):532-553. doi:10.1159/000485089
- Zhao Y, Scott NA, Fynch S, et al. Autoreactive T cells induce necrosis and not BCL-2-regulated or death receptor-mediated apoptosis or RIPK3-dependent necroptosis of transplanted islets in a mouse model of type 1 diabetes. *Diabetologia*. 2015;58(1):140-148.
- Jörns A, Amdt T, Zu Vilsendorf AM, et al. Islet infiltration, cytokine expression and beta cell death in the NOD mouse, BB rat, Komeda rat, LEW.1AR1-iddm rat and humans with type 1 diabetes. *Diabetologia*. 2014;57(3):512-521. doi:10.1007/s00125-013-3125-4
- Wilcox NS, Rui J, Hebrok M, Herold KC. Life and death of β cells in Type 1 diabetes: A comprehensive review. J Autoimmun. 2016; 71:51-58. doi:10.1016/j.jaut.2016.02.001
- 24. Schieber M, Chandel NS. ROS function in redox signaling. Curr Biol. 2014;24(10):453-462. doi:10.1016/j.cub.2014.03.034.ROS
- Kikumoto Y, Sugiyama H, Inoue T, et al. Sensitization to alloxan-induced diabetes and pancreatic cell apoptosis in acatalasemic mice. Biochim Biophys Acta - Mol Basis Dis. 2010;1802(2):240-246. doi:10.1016/j.bbadis.2009.10.009
- Kannan K, Jain SK. Oxidative stress and apoptosis. Pathophysiology. 2000;7(27):153-163. doi:10.1016/s0928-4680(00)00053-5
- Sinha K, Das J, Pal PB, Sil PC. Oxidative stress: The mitochondria-dependent and mitochondria-independent pathways of apoptosis. Arch Toxicol. 2013;87(7):1157-1180. doi:10.1007/s00204-013-1034-4
- Marroqui L, Dos Santos RS, Fløyel T, et al. TYK2, a candidate gene for type 1 diabetes, modulates apoptosis and the innate immune response in human pancreatic β-cells. Diabetes. 2015;64(11):3808-3817. doi:10.2337/db15-0362
- Yamamoto M, Taniguchi S, Aoyagi K. Domain structure as affected by the uniaxial ferromagnetic anisotropy induced in cubic solid solutions. *Phys Rev.* 1956;102(5):1295-1297. doi:10.1103/PhysRev.102.1295
- 30. Prentki M, Nolan CJ. Islet β cell failure in type 2 diabetes. J Clin Invest. 2006;116(7):1802-1812. doi:10.1172/JCI29103
- Gonçalves RV, Costa AMA, Grzeskowiak L. Oxidative stress and tissue repair: mechanism, biomarkers, and therapeutics. Oxid Med Cell Longev. 2021; 2021:12-14. doi:10.1155/2021/6204096
- 32. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr Sci. 2016 Dec 29;5: e47):1-15. doi:10.1017/jns.2016.41
- Sailaja Rao P, Kalva S, Yerramilli A, Mamidi S. Free radicals and tissue damage: role of antioxidants. Free Radicals Antioxidants. 2011;1(4):1-6. doi:10.5530/ax.2011.4.2
- Beecher GR. Introduction to the Proceedings of the Third International Scientific Symposium on Tea and Human Health: Role of flavonoids in the diet. J Nutr. 2003;133(10):3248-3254. doi:10.1093/jn/133.10.3248S
- Gothai S, Ganesan P, Park S-Y, Fakurazi S, Choi D-K, Arulselvan P. Natural phyto-bioactive compounds for the treatment of type 2 diabetes: inflammation as a target. Nutrients. 2016;8(8):461. doi:10.3390/nu8080461
- Al-Ishaq RK, Abotaleb M, Kubatka P, Kajo K, Büsselberg D. Flavonoids and their anti-diabetic effects: Cellular mechanisms and effects to improve blood sugar levels. Biomolecules. 2019;9(9):1-35. doi:10.3390/biom9090430
- Lee MS, Chyau CC, Wang CP, Wang TH, Chen JH, Lin HH. Flavonoids identification and pancreatic beta-cell protective effect of lotus seedpod. Antioxidants. 2020;9(8):1-23. doi:10.3390/antiox9080658
- Laddha AP, Kulkarni YA. Tannins and vascular complications of Diabetes: An update. Phytomedicine. 2019; 56:229-245. doi:10.1016/j.phymed.2018.10.026
- Ali Asgar M. Anti-diabetic potential of phenolic compounds: A review. Int J Food Prop. 2013;16(1):91-103. doi:10.1080/10942912.2011.595864
- Tong WY, Wang H, Waisundara VY, Huang D. Inhibiting enzymatic starch digestion by hydrolyzable tannins isolated from Eugenia jambolana. LWT - Food Sci Technol. 2014;59(1):389-395.
- Matsui T, Ueda T, Oki T, Sugita K, Terahara N, Matsumoto K. α-glucosidase inhibitory action of natural acylated anthocyanins. 1. Survey of natural pigments with potent inhibitory activity. J Agric Food Chem. 2001;49(4):1948-1951. doi:10.1021/jf001251u
- Amiraragab B, Hussein SA, Alm-Eldeen A-E, Hafe z A, Mohamed T. Diabetes management saponins and their potential role in diabetes mellitus. *Diabetes Manag*. 2017;7(1):148-158.
- Elekofehinti OO. Saponins: Anti-diabetic principles from medicinal plants A review. Pathophysiology. 2015;22(2):95-103. doi:10.1016/i.pathophys.2015.02.001
- 44. Ashour AS, El Aziz MMA, Gomha Melad AS. A review on saponins from medicinal plants: chemistry, isolation, and determination. J

- Nanomedicine Res. 2019;7(4):282-288. doi:10.15406/jnmr.2019.07.00199
- Sharma B, Mittal A, Dabur R. Mechanistic approach of anti-diabetic compounds identified from natural sources. Chem Biol Lett. 2018;5(2):63-99.
- Nafiu M, Tom Ashafa A. Antioxidant and inhibitory effects of saponin extracts from Dianthus basuticus Burtt Davy on key enzymes implicated in type 2 diabetes In vitro. *Pharmacogn Mag*. 2017;13(52):576-582. doi:10.4103/pm.pm\_583\_16
- Kumar A, Aswal S, Semwal RB, Chauhan A, Joshi SK, Semwal DK. Role of plant-derived alkaloids against diabetes and diabetes-related complications: a mechanism-based approach. *Phytochem Rev.* 2019;18(5):1277-1298. doi:10.1007/s11101-019-09648-6
- Larantukan SVM, Setiasih LNE, Widyastuti SK, et al. Pemberian Ekstrak Etanol Kulit Batang Kelor Glukosa Darah Tikus Hiperglikemia. Indones Med Veterinus. 2014;3(4):292-299.
- Unuofin JO, Lebelo SL. Antioxidant effects and mechanisms of medicinal plants and their bioactive compounds for the prevention and treatment of type 2 diabetes: An Updated Review. Oxid Med Cell Longev. 2020; 2020:1-36. doi:10.1155/2020/1356893
- Putta S, Sastry Yarla N, Kumar Kilari E, et al. Therapeutic potentials of triterpenes in diabetes and its associated complications. Curr Top Med Chem. 2016;16(23):2532-2542. doi:10.2174/1568026616666160414123343
- Lyu H, Chen J, Li WL. Natural triterpenoids for the treatment of diabetes mellitus: A review. Nat Prod Commun. 2016;11(10):1579-1586. doi:10.1177/1934578x1601101037
- Nazaruk J, Borzym-Kluczyk M. The role of triterpenes in the management of diabetes mellitus and its complications. Phytochem Rev. 2015;14(4):675-690. doi:10.1007/s11101-014-9369-x
- Iskender H, Yenice G, Dokumacioglu E, Kaynar O, Hayirli A, Kaya A. The effects of dietary flavonoid supplementation on the antioxidant status of laying hens. Rev Bras Cienc Avic. 2016;18(4):663-668. doi:10.1590/1806-9061-2016-0356
- Ciampi F, Sordillo LM, Gandy JC, et al. Evaluation of natural plant extracts as antioxidants in a bovine in vitro model of oxidative stress. J Dairy Sci. 2020;103(10):8938-8947. doi:10.3168/jds.2020-18182
- Agati G, Brunetti C, Fini A, et al. Are flavonoids effective antioxidants in plants? Twenty years of our investigation. Antioxidants. 2020;9(11):1-17. doi:10.3390/antiox9111098
- Henneberg R, Otuki MF, Furman AEF, Hermann P, Nascimento AJ do, Leonart MSS. Protective effect of favonoids against reactive oxygen species production in sickle cell anemia patients treated with hydroxyurea. Rev Bras Hematol Hemoter. 2013;35(1):52-55. doi:10.5581/1516-8484.20130015
- Brunetti C, Di Ferdinando M, Fini A, Pollastri S, Tattini M. Flavonoids as Antioxidants and Developmental Regulators: Relative Significance in Plants and Humans. Int J Mol Sci. 2013;14(2):3540-3555. doi:10.3390/ijms14023540
- 58. Banjarnahor SDS, Artanti N. Antioxidant properties of flavonoids. Med J Indones. 2014;23(4):239-244. doi:10.13181/mji.v23i4.1015
- Xu D, Hu M-J, Wang Y-Q, Cui Y-L. Antioxidant Activities of Quercetin and Its Complexes for Medicinal Application. Molecules. 2019;24(6):1123. doi:10.3390/molecules24061123
- Amarowicz R. Tannins: the new natural antioxidants? Eur J Lipid Sci Technol. 2007;109(6):549-551. doi:10.1002/ejlt.200700145
- Sieniawska E. Activities of tannins-From in Vitro studies to clinical trials. Nat Prod Commun. 2015;10(11):1877-1884. doi:10.1177/1934578x1501001118
- Velayutham R, Sankaradoss N, Ahamed KN. Protective effect of tannins from Ficus racemosa in hypercholesterolemia and diabetes induced vascular tissue damage in rats. Asian Pac J Trop Med. 2012;5(5):367-373. doi:10.1016/S1995-7645(12)60061-3
- Ashraf MF, Abd Aziz M, Stanslas J, Ismail I, Abdul Kadir M. Assessment of antioxidant and cytotoxicity activities of saponin and crude extracts of Chlorophytum borivilianum. Sci World J. 2013; 2013;1-7. doi:10.1155/2013/216894
- Gülçin I, Mshvildadze V, Gepdiremen A, Elias R. Antioxidant activity of saponins isolated from ivy: α-Hederin, hederasaponin-C, hederacolchiside-E and hederacolchiside-F. Planta Med. 2004;70(6):561-563. doi:10.1055/s-2004-827158
- Lim JG, Park HM, Yoon KS. Analysis of saponin composition and comparison of the antioxidant activity of various parts of the quinoa plant (Chenopodium quinoa Willd.). Food Sci Nutr. 2020;8(1):694-702. doi:10.1002/fsn3.1358
- Chen Y, Miao Y, Huang L, et al. Antioxidant activities of saponins extracted from Radix Trichosanthis: An in vivo and in vitro evaluation. BMC Complement Altern Med. 2014;14(1):1-8. doi:10.1186/1472-6882-14-86
- 67. Muthuraman A, Krishan S, Perumal Ps, Anaswara P. Therapeutic potency of saponin rich aqueous extract of Scoparia dulcis L. in alloxan induced diabetes in rats. AYU (An Int Q J Res Ayurveda). 2014 Apr;35(2):211-7. doi:10.4103/0974-8520.146261
- Li YN, Guo Y, Xi MM, et al. Saponins from aralia taibaiensis attenuate D-galactose-induced aging in rats by activating FOXO3a and Nrf2 pathways. Oxid Med Cell Longev. 2014; 2014;320513. doi:10.1155/2014/320513
- El Guiche R, Tahrouch S, Amri O, El Mehrach K, Hatimie A. Antioxidant activity and total phenolic and flavonoid contents of 30 medicinal and aromatic plants located in the south of Morocco. Int J New Technol Res. 2015;1(3):7-11.
- Spiridon I, Bodirlau R, Teaca CA. Total phenolic content and antioxidant activity of plants used in traditional Romanian herbal medicine. Cent Eur J Biol. 2011;6(3):388-396. doi:10.2478/s11535-011-0028-6
- Pereira DM, Valentão P, Pereira JA, Andrade PB. Phenolics: From chemistry to biology. Molecules. 2009;14(6):2202-2211. doi:10.3390/molecules14062202
- Baharuddin NAF, Nordin MFM, Morad NA, Aris NIA, Yunus MAC. Total phenolic, flavonoid content and antioxidant activity of Clinacanthus nutans leaves by water-based ultrasonic assisted extraction. Malaysian J Anal Sci. 2018;22(4):659-666. doi:10.17576/mjas-2018-2204-12
- Augusto TR, Scheuermann Salinas ES, Alencar SM, D'Arce MABR, De Camargo AC, Vieira TMF de S. Phenolic compounds and antioxidant activity of hydroalcoholic extractsof wild and cultivated murtilla (Ugni molinae turcz.). Food Sci Technol. 2015;34(4):667-673. doi:10.1590/1678-457X.6393
- 74. Pourreza N. Phenolic compounds as potential antioxidant. Jundishapur J Nat Pharm Prod. 2013;8(4):149-150. doi:10.17795/jjnpp-15380
- Abdul-hafeez EY, Karamova N, Ilinskaya O. Antioxidant activity and total phenolic compound content of certain medicinal plants. Int J Biosci. 2014;5(9):213-222. doi:10.12692/ijb/5.9.213-222
- Gülçin I, Elias R, Gepdiremen A, Chea A, Topal F. Antioxidant activity of bisbenzylisoquinoline alkaloids from Stephania rotunda: Cepharanthine and fangchinoline. J Enzyme Inhib Med Chem. 2010;25(1):44-53. doi: 10.3109/14756360902932792
- Zahari A, Ablat A, Sivasothy Y, Mohamad J, Choudhary MI, Awang K. In vitro antiplasmodial and antioxidant activities of bisbenzylisoquinoline alkaloids from Alseodaphne comeri Kosterm. Asian Pac J Trop Med. 2016;9(4):328-332. doi:10.1016/j.apjtm.2016.03.008
- Hasanuzzaman M, Bhuyan MHMB, Zulfiqar F, et al. Reactive oxygen species and antioxidant defense in plants under abiotic stress: Revisiting the crucial role of a universal defense regulator. Antioxidants. 2020;9(8):1-52. doi:10.3390/antiox9080681

- Kooti W, Farokhipour M, Asadzadeh Z, Ashtary-Larky D, Asadi-Samani M. The role of medicinal plants in the treatment of diabetes: a systematic review. Electron physician. 2016;8(1):1832-1842. doi:10.19082/1832
- Forni C, Facchiano F, Bartoli M, et al. Beneficial role of phytochemicals on oxidative stress and age-related diseases. Biomed Res Int. 2019 Apr 7; 2019:8748253. doi:10.1155/2019/8748253
- Tiong SH, Looi CY, Hazni H, et al. Antidiabetic and antioxidant properties of alkaloids from Catharanthus roseus (L.) G. Don. Molecules. 2013;18(8):9770-9784. doi:10.3390/molecules18089770
- 82. Kasote DM, Katyare SS, Hegde M V., Bae H. Significance of antioxidant potential of plants and its relevance to therapeutic applications. Int J Biol Sci. 2015;11(8):982-991. doi:10.7150/ijbs.12096
- 83. Gülçin I, Mshvildadze V, Gepdiremen A, Elias R. The antioxidant activity of a triterpenoid glycoside isolated from the berries of Hedera colchica: 3-O-(β-D-glucopyranosyl)-hederagenin. Phyther Res. 2006;20(2):130-134. doi:10.1002/ptr.1821
- Xi M, Hai C, Tang H, et al. Antioxidant and antiglycation properties of triterpenoid saponins from Aralia taibaiensis traditionally used for treating diabetes mellitus. Redox Rep. 2010;15(1):20-28. doi:10.1179/174329210X12650506623041
- Nzogong RT, Ndjateu FST, Ekom SE, et al. Antimicrobial and antioxidant activities of triterpenoid and phenolic derivatives from two Cameroonian Melastomataceae plants: Dissotis senegambiensis and Amphiblemma monticola. BMC Complement Altern Med. 2018;18(1):1-12. doi:10.1186/s12906-018-2229-2
- Liu CH, Yen MH, Tsang SF, Gan KH, Hsu HY, Lin CN. Antioxidant triterpenoids from the stems of Momordica charantia. Food Chem. 2010;118(3):751-756. doi:10.1016/j.foodchem.2009.05.058
- 87. Zhu L, Yi X, Ma C, et al. Betulinic Acid Attenuates Oxidative Stress in the Thymus Induced by Acute Exposure to T-2 Toxin via Regulation of the MAPK/Nrf2 Signaling Pathway. Toxins (Basel). 2020;12(9):540. doi:10.3390/toxins12090540
- Ahmad R, Khan A, Lee HJ, et al. Lupeol, a plant-derived triterpenoid, protects mice brains against Aβ-induced oxidative stress and neurodegeneration. Biomedicines . 2020;8(10):380. doi:10.3390/biomedicines8100380
- Teng H, Yuan B, Gothai S, Arulselvan P, Song X, Chen L. Dietary triterpenes in the treatment of type 2 diabetes: To date. Trends Food Sci Technol. 2018;72 (June 2017):34-44. doi:10.1016/j.tifs.2017.11.012
- Adewole SO, Caxton-Martins EA, Ojewole JAO. Protective effect of quercetin on the morphology of pancreatic β-cells of stretozotocintreated diabetic rats. Afr J Tradit Complement Altern Med. 2007;4(1):64-74. doi:10.4314/ajtcam.v4i1.31196
- Fetouh FA, Azab AES. Ameliorating effects of Curcumin and Propolis against the reproductive toxicity of gentamicin in adult male guinea pigs: quantitative analysis and morphological study. Am J Life Sci. 2014;2(3):138-149. doi:10.11648/j.ajls.20140203.13
- Adwas AA, Ibrahim ASE, Azab AE, Quwaydir FA. Oxidative stress and antioxidant mechanisms in human body. J Appl Biotechnol Bioeng. 2019;6(1):43-47. doi:10.15406/jabb.2019.06.00173
- 93. Jiménez-Osorio AS, González-Reyes S, Pedraza-Chaverri J. Natural Nrf2 Activators in Diabetes. Clinica Chimica Acta. 2015 Aug 25; 448:182-92. doi:10.1016/j.cca.2015.07.009
- 94. Wang YJ, Wang XY, Hao XY, et al. Ethanol extract of centipeda minima exerts antioxidant and neuroprotective effects via activation of the Nrf2 signaling pathway. Oxid Med Cell Longev. 2019; 2019:1-16. doi:10.1155/2019/9421037
- Wang G, Fu Y, Li J, et al. Aqueous extract of Polygonatum sibiricum ameliorates ethanol-induced mice liver injury via regulation of the Nrf2/ARE pathway. J Food Biochem. 2021;45(1):1-11. doi:10.1111/jfbc.13537
- Jayawardena TU, Sanjeewa KKA, Fernando IPS, et al. Sargassum horneri (Tumer) C. Agardh ethanol extract inhibits the fine dust inflammation response via activating Nrf2/HO-1 signaling in RAW 264.7 cells. BMC Complement Altern Med. 2018;18(1):1-11. doi:10.1186/s12906-018-2314-6
- He F, Ru X, Wen T. NRF2, a transcription factor for stress response and beyond. Int J Mol Sci. 2020;21(13):1-23. doi:10.3390/ijms21134777
- Reis AA da S, Santos R da S, Cruz AH da S, Silva EG da, Cruz AD da, Pedrino GR. The Effect of Nrf2 on Diabetic Complications. In: A Master Regulator of Oxidative Stress - The Transcription Factor Nrf2.; 2016:131-144. doi:10.5772/66132
- Matveyenko A V., Butler PC. Relationship between β-cell mass and diabetes onset. Diabetes Obes Metab. 2008;10(4):23-31. doi:10.1111/j.1463-1326.2008.00939.x
- Cleaver O. β Cell Renewal versus Differentiation: Slow and Steady Wins the Race. Dev Cell. 2017;41(3):223-225. doi:10.1016/j.devcel.2017.04.017
- Porat S, Weinberg-Corem N, Tornovsky-Babaey S, et al. Control of pancreatic β cell regeneration by glucose metabolism. Cell Metab. 2011;13(4):440-449. doi:10.1016/j.cmet.2011.02.012
- 102. Bouwens L, Rooman I. Regulation of pancreatic beta-cell mass. Physiol Rev. 2005;85(4):1255-1270. doi:10.1152/physrev.00025.2004

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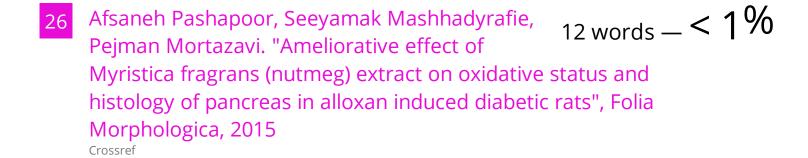
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- V. P. Lebedev, S. V. Bilichenko, A. V. Malygin, S. P. Nechiporenko, S. E. Kolbasov, M. V. Melikhova.

  "Transcranial Stimulation Normalizes Blood Sugar Levels in Alloxan Diabetes in Rats", Neuroscience and Behavioral Physiology, 2006

  Crossref
- Wilcox, Nicholas S., Jinxiu Rui, Matthias Hebrok, and Kevan C. Herold. "Life and death of  $\beta$  cells in Type 1 diabetes: A comprehensive review", Journal of Autoimmunity, 2016.
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- Tariq G. Fellous. "Cellular pathways to β-cell replacement", Diabetes/Metabolism Research and Reviews, 02/2007

  Crossref
- Yeni Susanti, Merryana Adriani, Annis Catur Adi. "Effect of Single Clove Garlic Extract (Allium Sativum Linn) on Blood Sugar Levels, Malondialdehyde, Insulin Levels and Insulin Resistance (Experiments in Rats (Rattus Novergicus) Induced by Streptozotocin", STRADA Jurnal Ilmiah Kesehatan, 2020

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- Tiffany Dennis, Michael Fanous, Shaker Mousa.

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  Crossref
- Yi-Jie Wang, Xin-Yue Wang, Xu-Yi Hao, Yong-Ming Yan et al. " Ethanol Extract of Exerts Antioxidant and Neuroprotective Effects via Activation of the Nrf2 Signaling Pathway ", Oxidative Medicine and Cellular Longevity, 2019 Crossref

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